SLiCE Reaction

1) Prepare 10X SLiCE Buffer in a 1.5 mL tube :

 $500 \ \mu L \ 1 \ M \ Tris-HCl \ pH \ 7.5$ + 50 \ \mu L \ 2 \ M \ MgCl_2 + 100 \ \mu L \ 100 \ mM \ ATP + 10 \ \mu L \ 1 \ M \ DTT + ____ddH_20 \ to 1 \ mL

Store at -20 C in 40-60 µl aliquots.

2) Add the following ingredients into a 0.2 mL tube in this order and **vortex**:

	linearized vector backbone (50-200 ng)
+	each additional assembly piece (1:1-1:10 molar ratio of
	vector:insert)
+1	μL 10X SLiCE buffer
+1	μL PPY SLiCE extract
+	ddH ₂ 0 to
10	μL

3) Incubate the SLICE reaction mix as above at 37 C for 1 hour using a PCR machine or water bath, and then place on ice.

4) Transform 1 μ L of the assembly reaction into 100 μ L of competent *E. coli* and/or run a diagnostic agarose gel to check for successful assembly.

5) For electroporation, transform 1 μ L into 20 μ L electrocompetent cells. For large recombinant DNA, electroporation is required. In complex cloning, electroporation is recommended, as it is 10-100 times as efficient as chemical transformation.